

# SMALL-MOLECULE MODULATION OF HUMAN NAPE-PLD SIGNALING

**Nanosciences – 3<sup>rd</sup> year PhD annual exam**

**Sara Chiarugi**

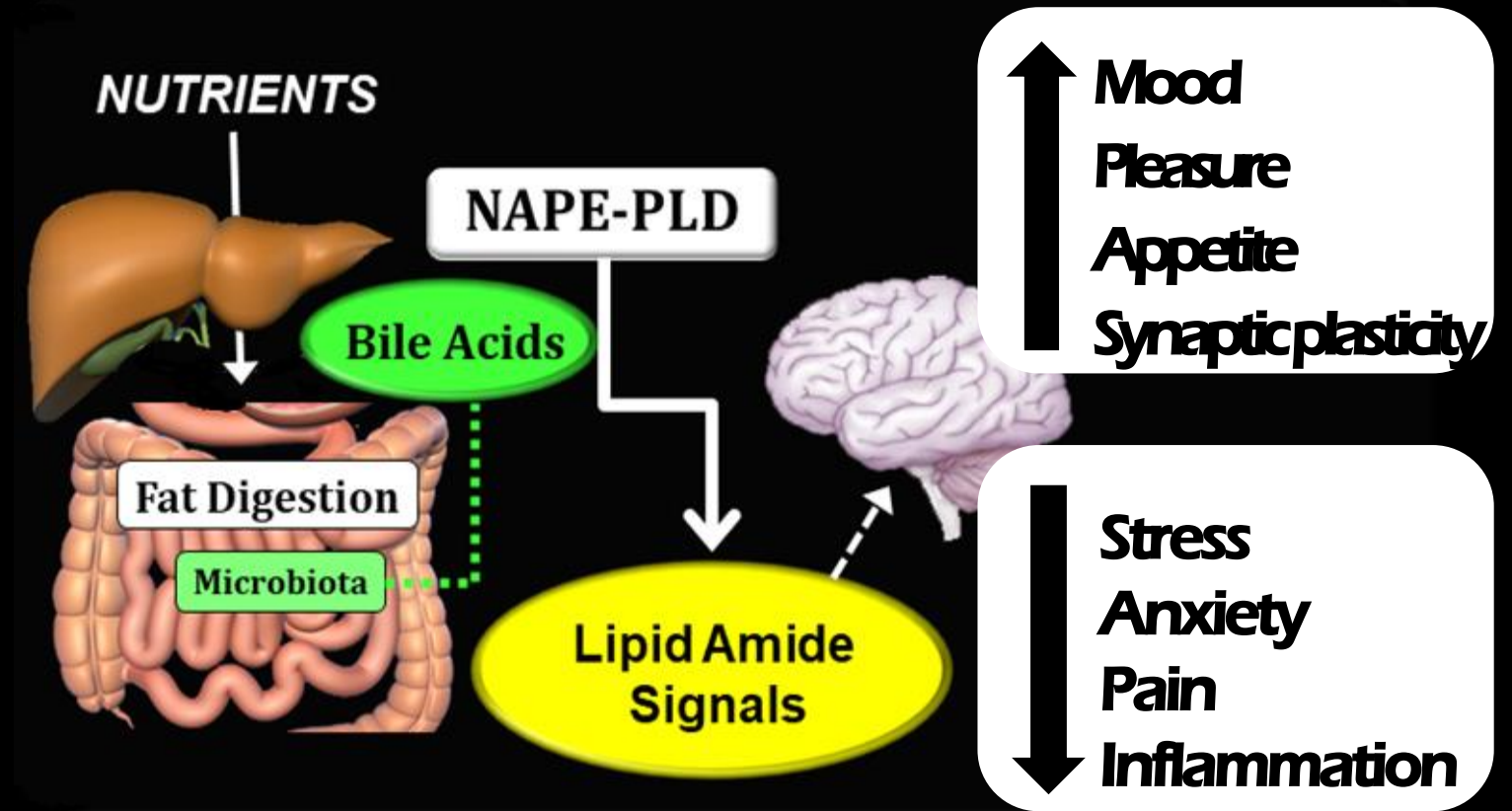
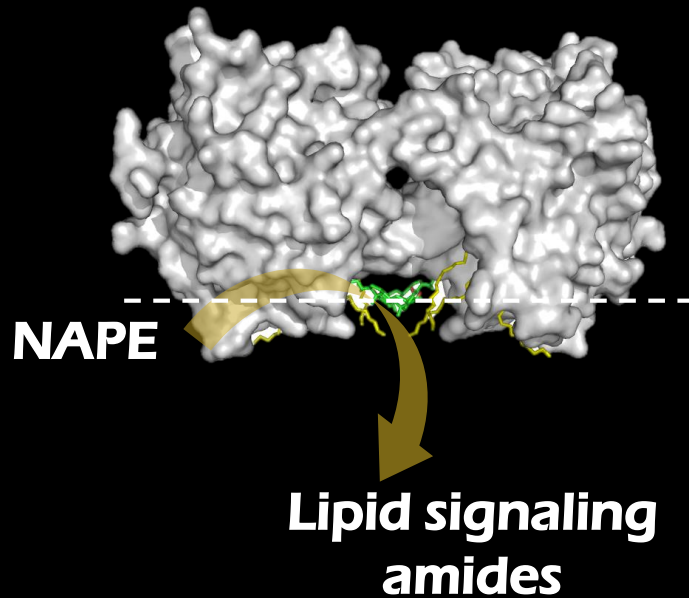
PhD student, Scuola Normale Superiore – Pisa

BIOSTRUCTURES Lab Dr. Gianpiero Garau,

ISTITUTO ITALIANO DI TECNOLOGIA, IIT@NEST – Pisa.

Pisa, 20/10/2020

# BIOACTIVE LIPID AMIDES

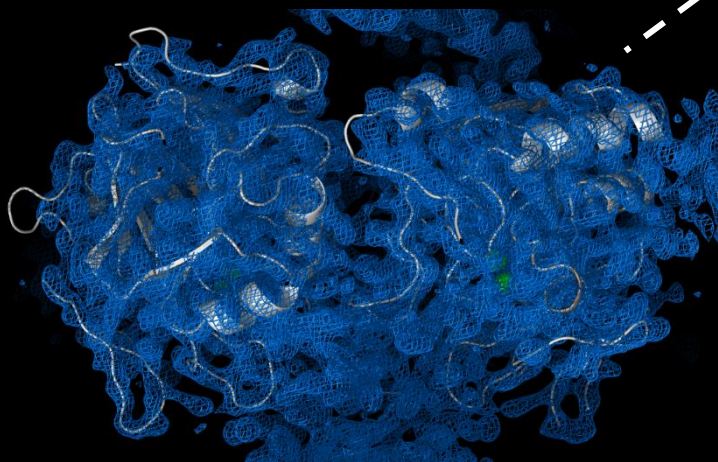
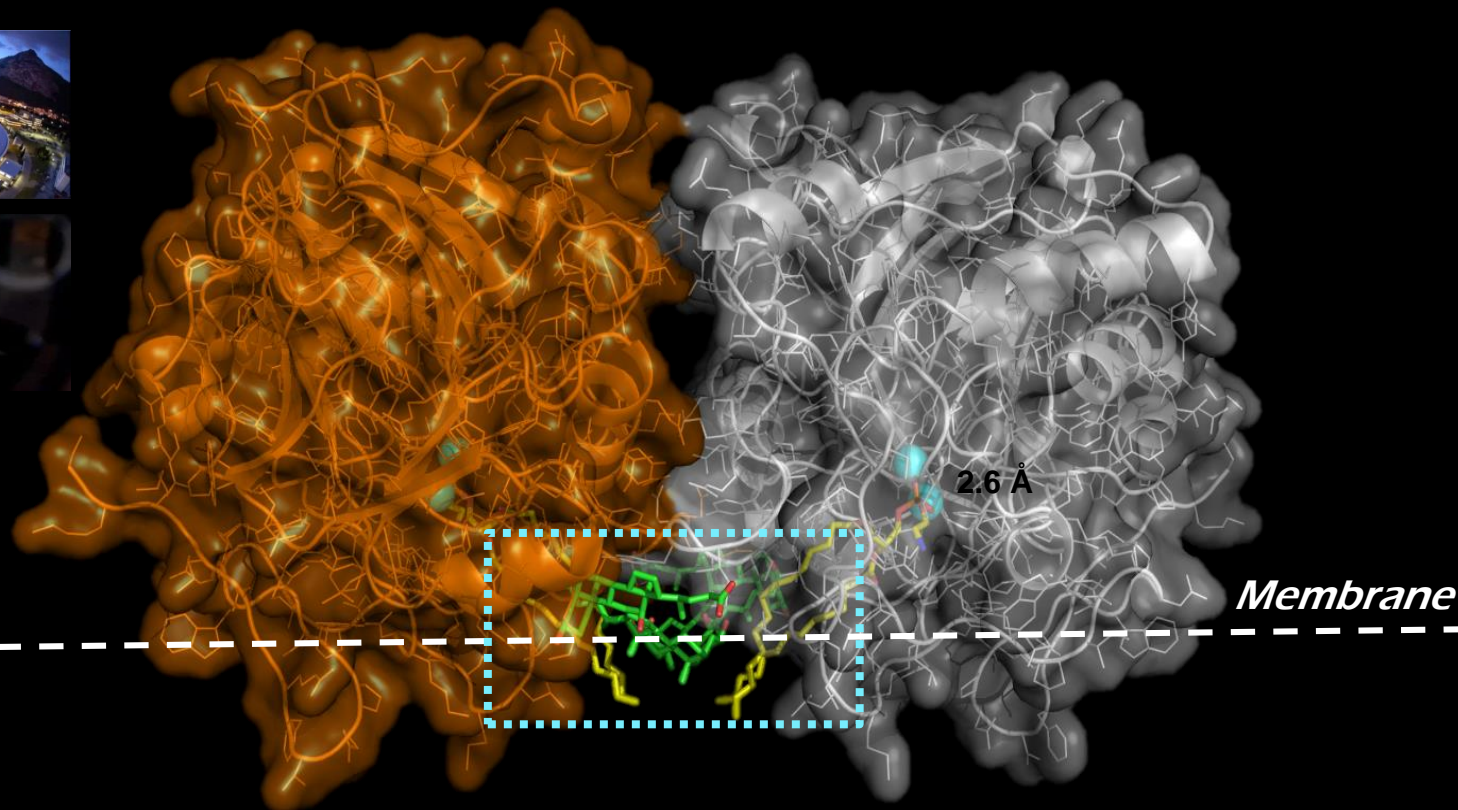


Modulation for potential therapeutic application

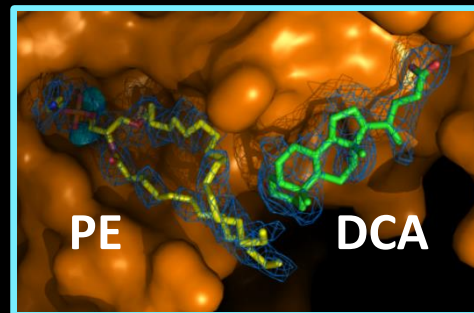
# STRUCTURE OF HUMAN MEMBRANE NAPE-PLD



**ELETTRA XRD1**  
**ESRF beamline ID23-2**

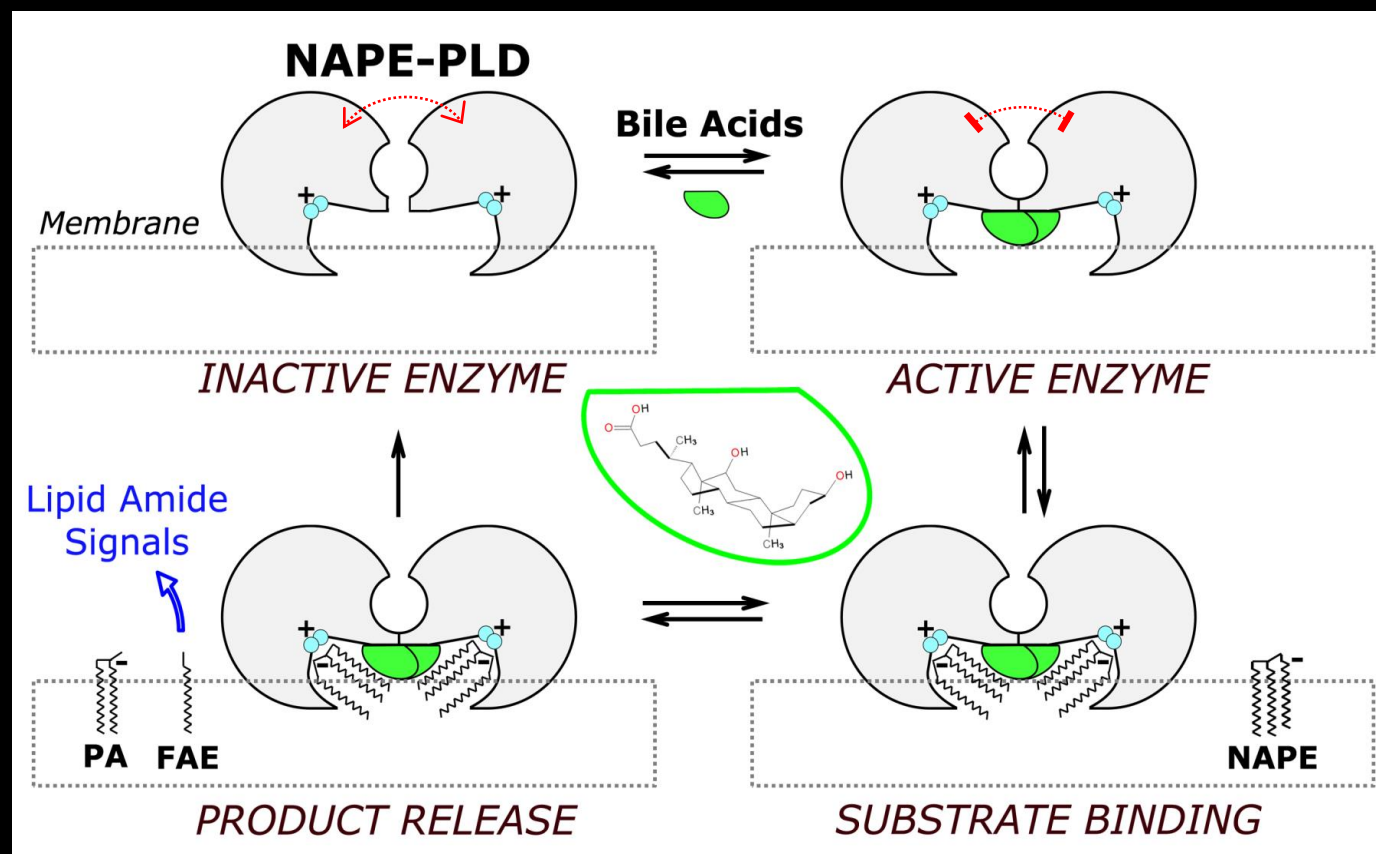


Structure 2015





# WHAT IS THE ROLE OF BAs?



**Dimer stabilization**  
**Enzyme activity**

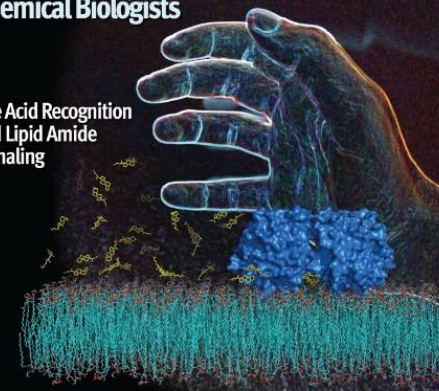
**Bile Acids turn  
NAPE-PLD on**

**ACS  
chemical  
biology**

OCTOBER 2016  
VOLUME 11, NUMBER 10  
pubs.acs.org/acchemicalbiology

The Community of  
Chemical Biologists

Bile Acid Recognition  
and Lipid Amide  
Signaling



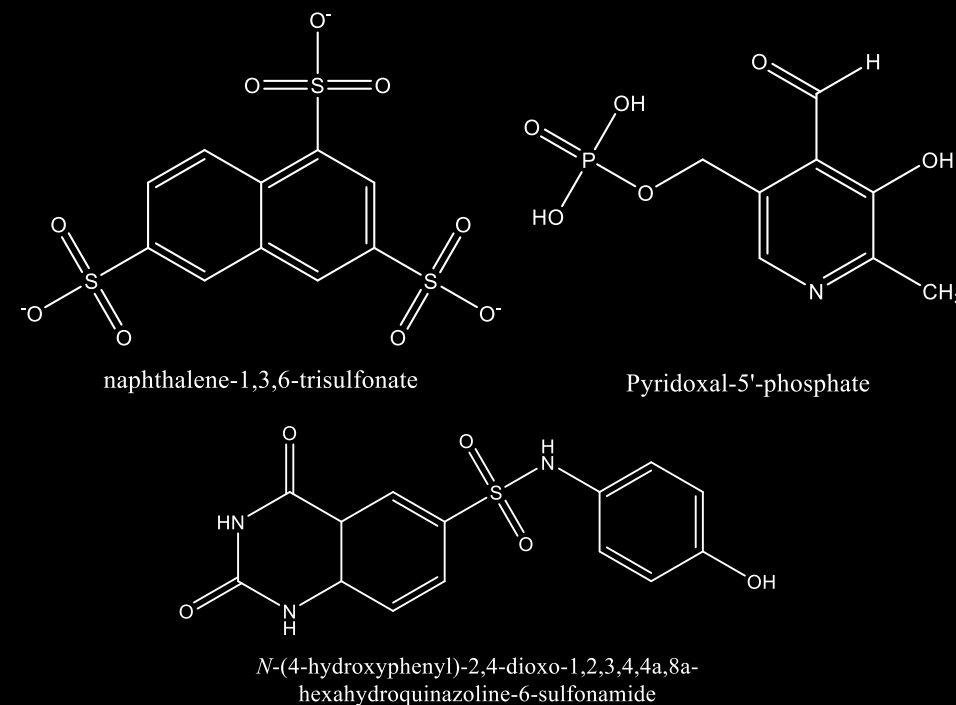
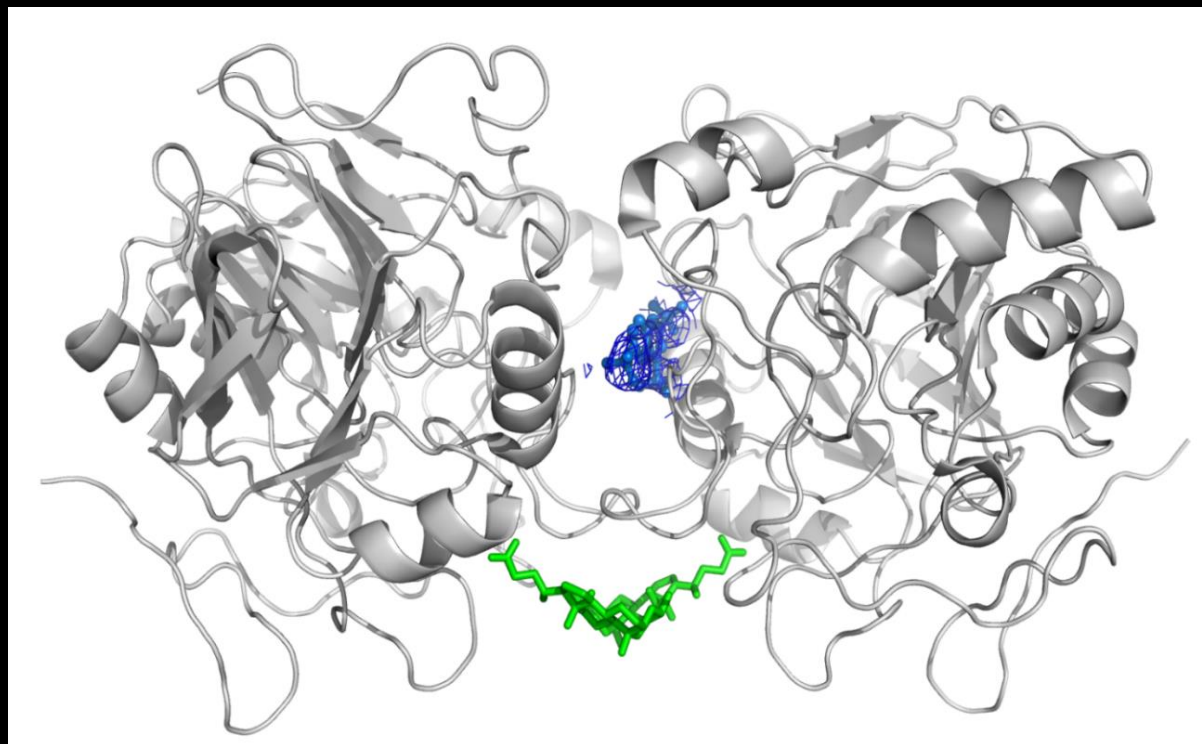
# LIGAND DISCOVERY

Ligand screening

SPR

Different Active  
Classes of Cmps

FRAGMENTS



First structures of NAPE-PLD in complex with chemical modulators

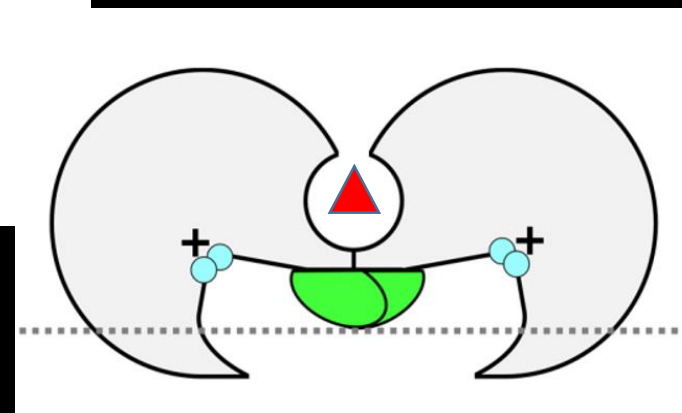
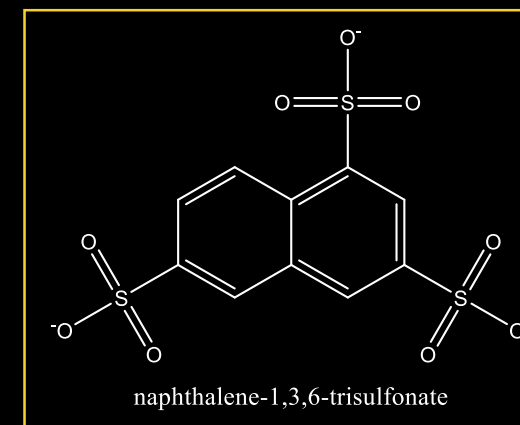
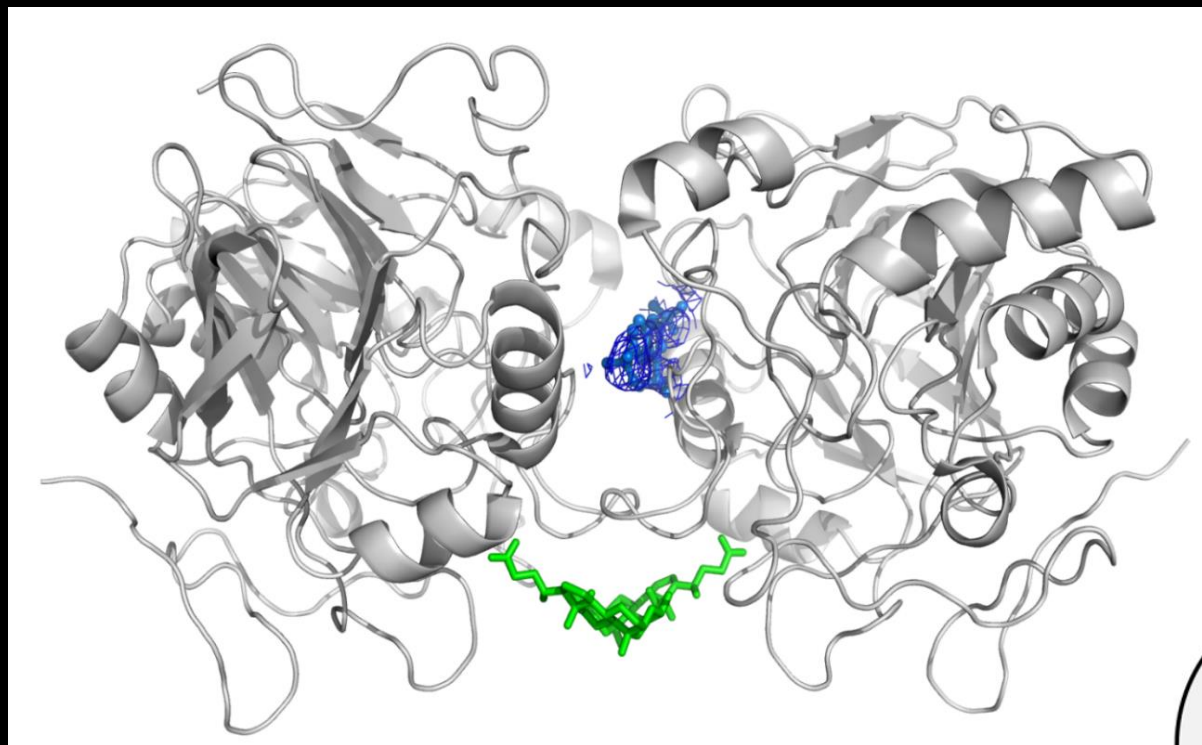
# Structure of NAPE-PLD in complex with Naph3S

Ligand screening

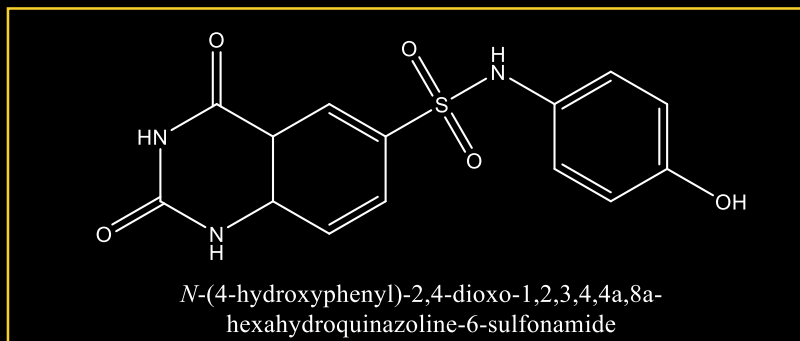
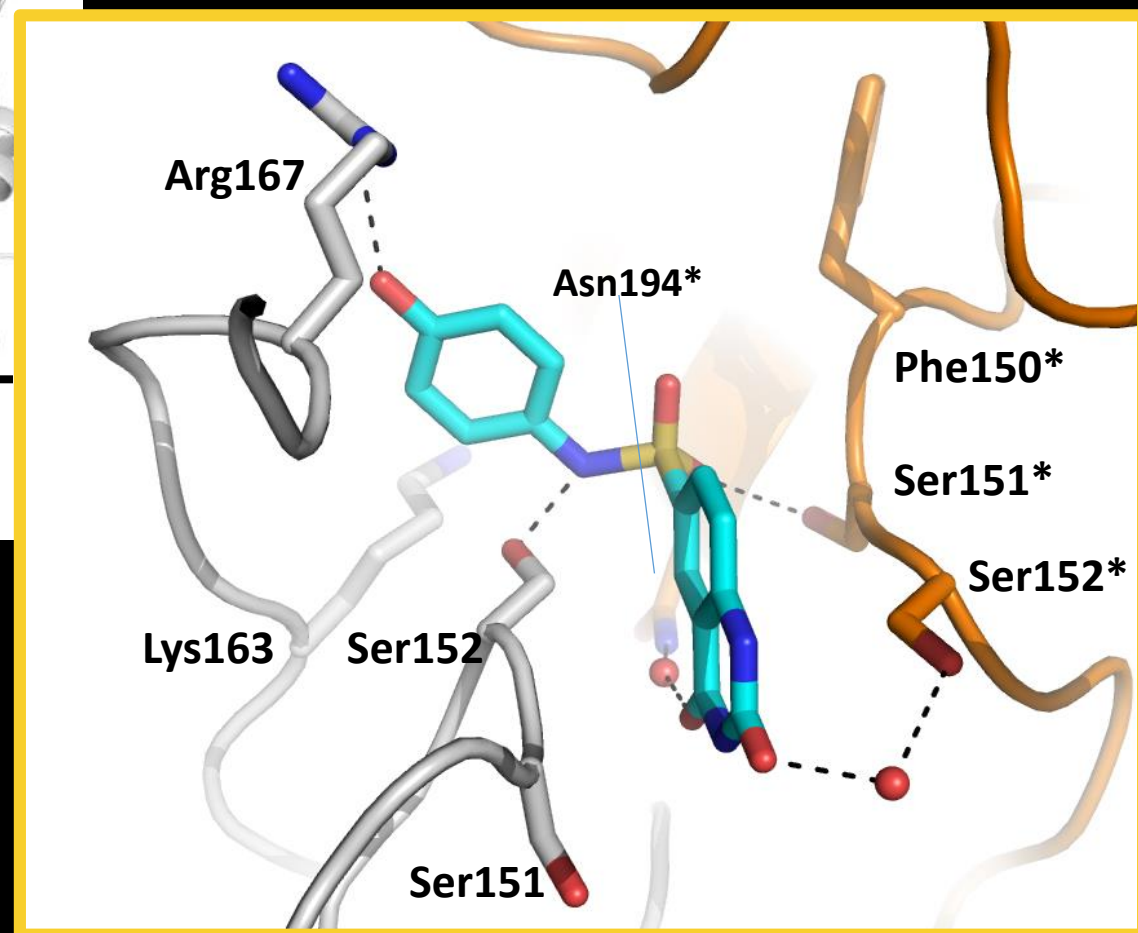
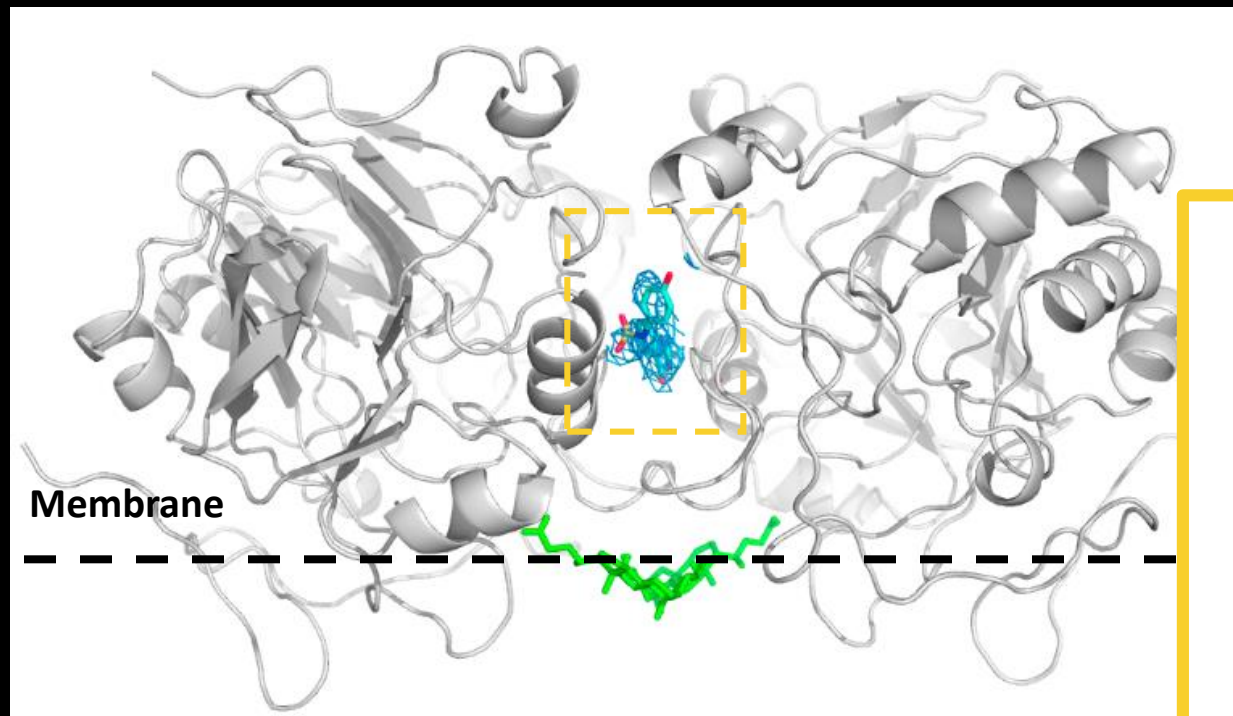
SPR

Different Active  
Classes of Cmps

FRAGMENTS



# Structure of NAPE-PLD in complex with ARN compound





# LIGAND DISCOVERY



## HHS Public Access

Author manuscript

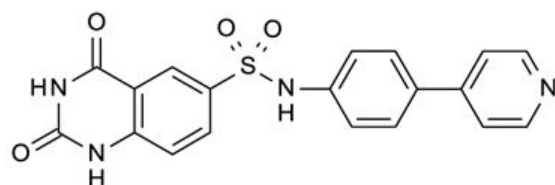
*Chem Commun (Camb)*. Author manuscript; available in PMC 2018 November 28.

Published in final edited form as:

*Chem Commun (Camb)*. 2017 November 28; 53(95): 12814–12817. doi:10.1039/c7cc07582k.

### Synthesis and characterization of the first inhibitor of N-acylphosphatidylethanolamine phospholipase D (NAPE-PLD)

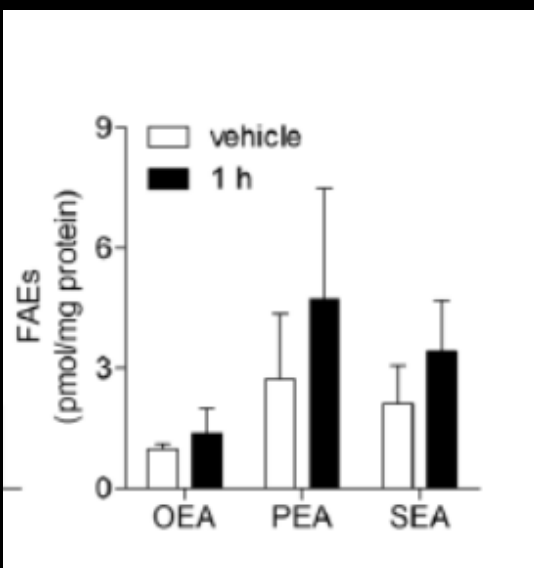
Beatrice Castellani<sup>a,±</sup>, Eleonora Diamanti<sup>a,#,±</sup>, Daniela Pizzirani<sup>a,‡</sup>, Piero Tardia<sup>a</sup>, Martina Maccesi<sup>c</sup>, Natalia Realini<sup>a</sup>, Paola Magotti<sup>a,&</sup>, Gianpiero Garau<sup>a,†</sup>, Thomas Bakkum<sup>a,§</sup>, Silvia Rivara<sup>d</sup>, Marco Mor<sup>d</sup>, and Daniele Piomelli<sup>\*,a,b</sup>



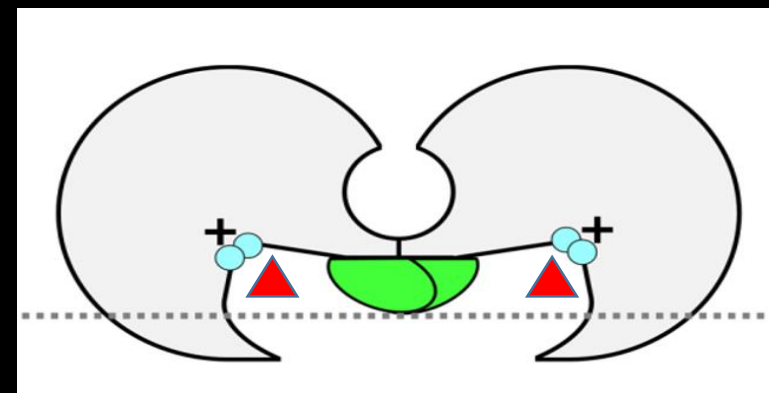
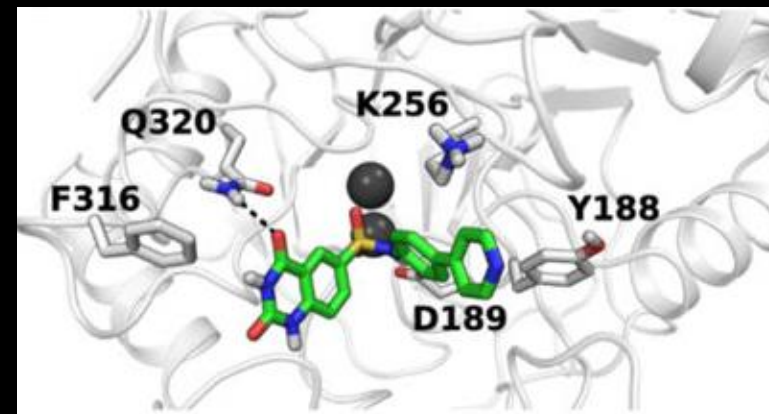
**ARN19874**

IC<sub>50</sub> ≈ 34 μM

NAPE-PLD inhibition (HEK)



In silico model

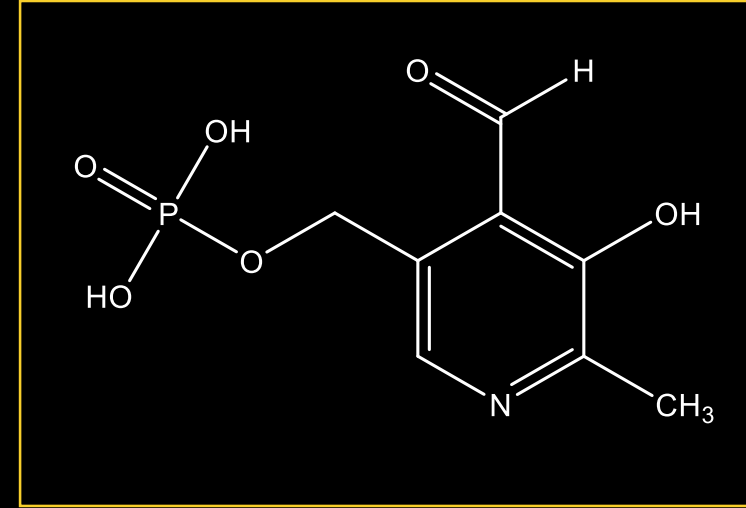
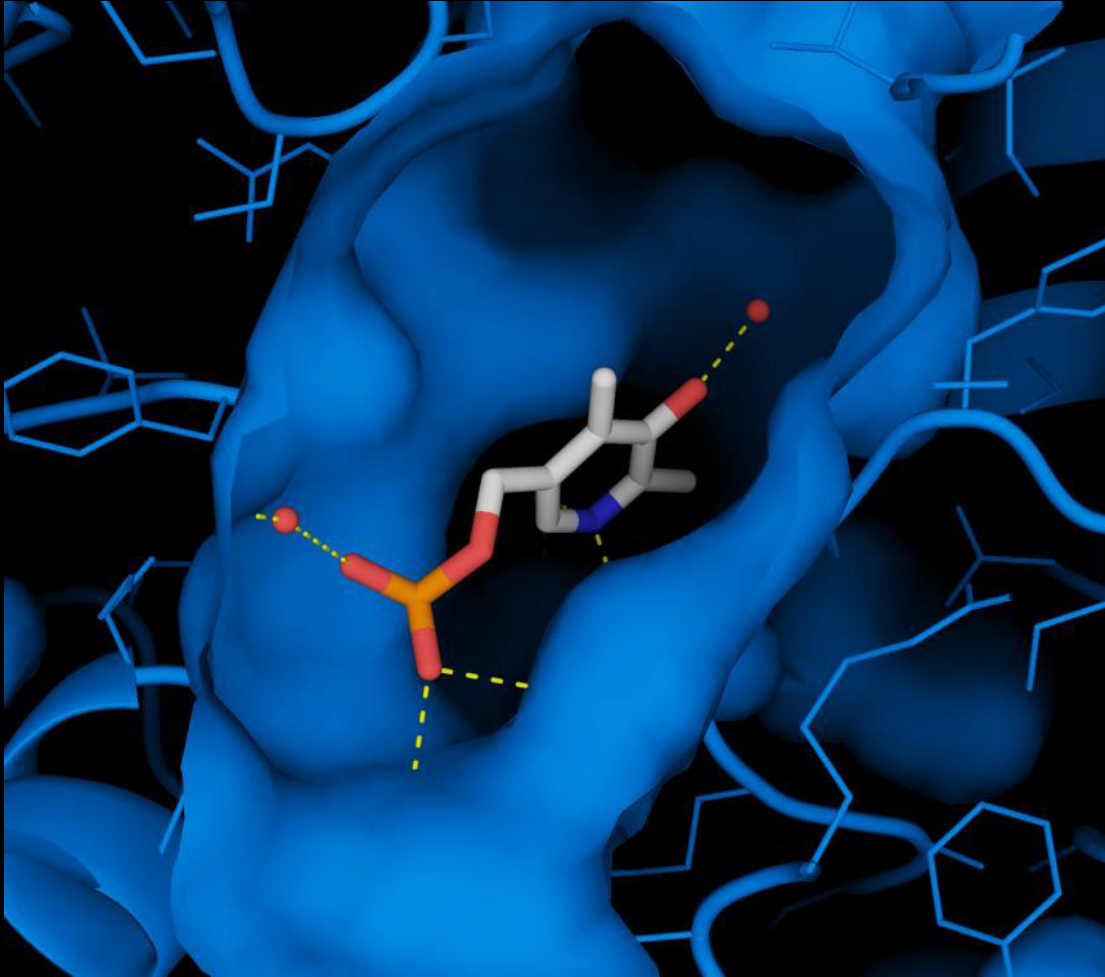


**VALIDATION by XRD**



# Structure of NAPE-PLD in complex with PLP

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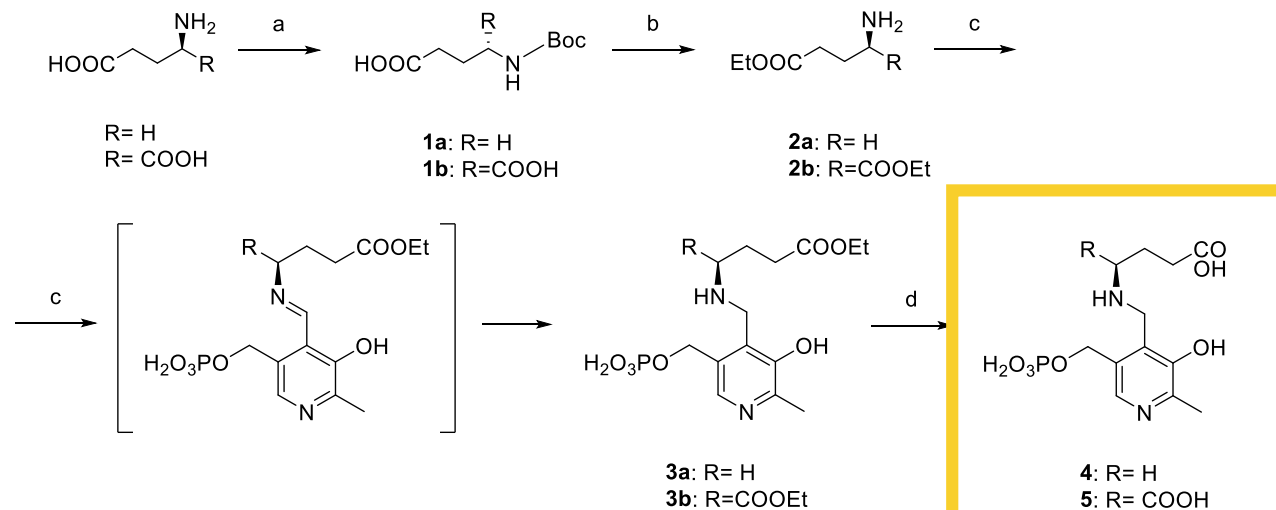


## Pyridoxal-5'-phosphate

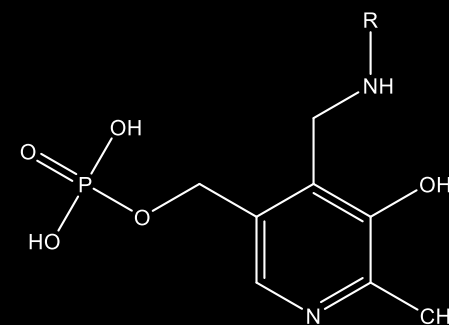
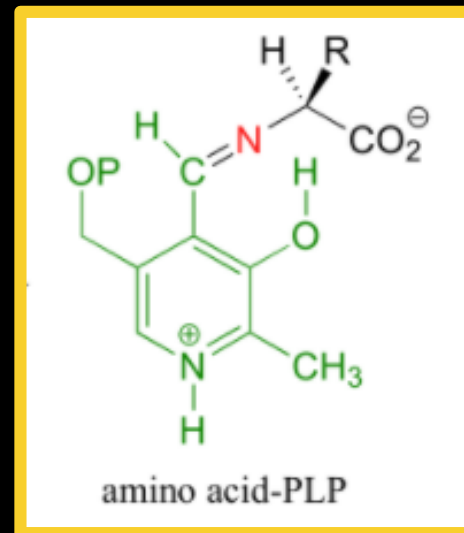
- ☐ Active form of Vitamin B6
- ☐ Involved in neurotransmitters generation
- ☐ lipid metabolism and gene expression

# PLP ANALOGUES

**SCHEME 1**

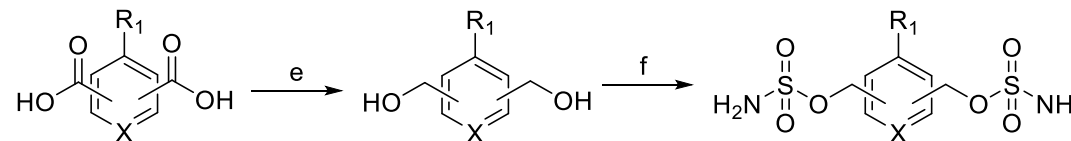


**Reagents and conditions:** a.  $\text{N}(\text{Et})_3$ , Di-tert-butyl dicarbonate,  $\text{DMF}/\text{H}_2\text{O}$ , 12 h; b.  $\text{EtOH}$ ,  $\text{SOCl}_2$ , r.t., 72 h; c. **1**) pyridoxal-5'-phosphate,  $\text{MeOH}$ , 30 min; **2**)  $\text{NaBH}_4$ ,  $5^\circ\text{C}$ , 15 min; **3**)  $25^\circ\text{C}$ , 72 h d.  $\text{EtOH}$ ,  $\text{NaOH}$  10%,  $69^\circ\text{C}$ , 12 h.



Prof. Simona Rapposelli  
University of Pisa  
Department of Pharmacy

**SCHEME 2**



**Reagents and conditions:** e.,  $\text{LiAlH}_4$ , dry  $\text{THF}$ , 20 h,  $0^\circ\text{C}$ ; f. sulfamoyl chloride,  $\text{DMA}$ , r.t.

# CONCLUSIONS & PERSPECTIVES

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**Affinity/kinetic of PLP-  
derivative**



**Dr. Aldo  
Moscardini**

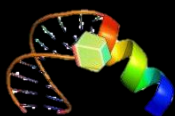
**Characterization of  
the structural complex  
by XRD**

**Modulation in cell  
and in vivo**



**Universiteit  
Leiden**

**Prof. Mario van der Stelt**



**GeCrySchool**



**"Insights into human membrane NAPE-PLD interactions" (manuscript in preparation)**



Universiteit  
Leiden

*Leading Discovery*

Prof. Mario van der Stelt

nature  
chemical biology

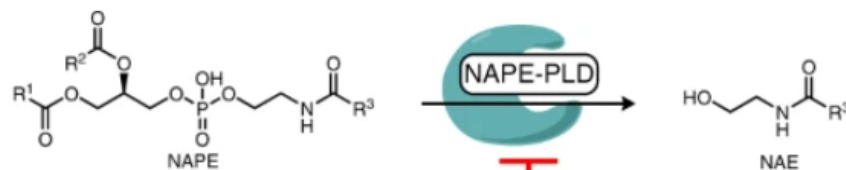
ARTICLES

<https://doi.org/10.1038/s41589-020-0528-7>

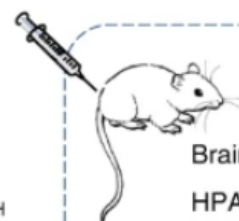
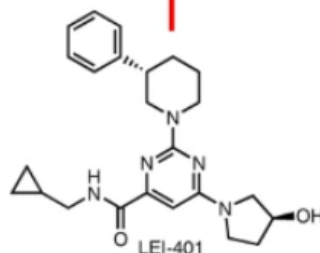
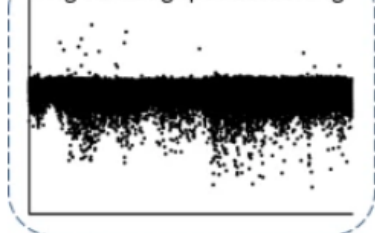


## Discovery of a NAPE-PLD inhibitor that modulates emotional behavior in mice

Elliot D. Mock<sup>1</sup>, Mohammed Mustafa<sup>2</sup>, Ozge Gunduz-Cinar<sup>3</sup>, Resat Cinar<sup>4</sup>, Gavin N. Petrie<sup>5</sup>, Vasudev Kantae<sup>1,6</sup>, Xinyu Di<sup>6</sup>, Daisuke Ogasawara<sup>7</sup>, Zoltan V. Varga<sup>8</sup>, Janos Paloczi<sup>8</sup>, Cristina Miliano<sup>9</sup>, Giulia Donvito<sup>2</sup>, Annelot C. M. van Esbroeck<sup>1</sup>, Anouk M. F. van der Gracht<sup>1</sup>, Ioli Kotsogianni<sup>1</sup>, Joshua K. Park<sup>4</sup>, Andrea Martella<sup>1</sup>, Tom van der Wel<sup>1,10</sup>, Marjolein Soethoudt<sup>1</sup>, Ming Jiang<sup>1,10</sup>, Tiemen J. Wendel<sup>1</sup>, Antonius P. A. Janssen<sup>1,10</sup>, Alexander T. Bakker<sup>1</sup>, Colleen M. Donovan<sup>3</sup>, Laura I. Castillo<sup>3</sup>, Bogdan I. Florea<sup>11</sup>, Jesse Wat<sup>12</sup>, Helma van den Hurk<sup>12</sup>, Matthias Wittwer<sup>13</sup>, Uwe Grether<sup>13</sup>, Andrew Holmes<sup>3</sup>, Constant A. A. van Boeckel<sup>1,12</sup>, Thomas Hankemeier<sup>6</sup>, Benjamin F. Cravatt<sup>7</sup>, Matthew W. Buczynski<sup>9</sup>, Matthew N. Hill<sup>5</sup>, Pal Pacher<sup>8</sup>, Aron H. Lichtman<sup>2,14</sup> and Mario van der Stelt<sup>1,10</sup> ✉



High-throughput screening



Brain NAEs ↓  
HPA axis ↑  
Fear extinction ↓